

PROGRAM PLANNING COMMITTEE

Jim Wells, Ph.D. (*Chair*) Professor and Chair, Dept of Pharmaceutical Chemistry University of California, San Francisco Oscar Millet, Ph.D Structural Biology Unit CIC bioGUNE Mark A. Lemmon, Ph.D. Professor and Chair, Dept of Biochemistry and Biophysics University of Pennsylvania Perelman School of Medicine

Miquel Pons, Ph.D.
Professor, Dept of Organic Chemistry
Universitat de Barcelona

Gary Pielak, Ph.D.
Distinguished Term Professor of Chemistry,
Biochemistry and Biophysics
University of North Carolina, Chapel Hill

TABLE OF CONTENTS

Symposium Overview	3
Executive Council & Leadership	4
Committees	5
Protein Society Awards	6
Call for 2015 Nominations	8
Hotel Floorplan	9
General Information	10
Travel Awards	12
Donor Acknowledgements	13
Daily Program	14
Day 1	14
Day 2	17
Day 3	21
Day 4	24
Poster Sessions	26
Exhibitors	47
Exhibitor Workshops	54
Speaker Abstracts	55
Poster Abstracts	
Author Index	274



Grand Hall A & B

Author Presentation Schedule:

Sunday, 07/27/2014 P02 - P08 Even Numbers 11:30 AM-01:30 PM P02 - P08 Odd Numbers 4:30 PM — 6:30 PM Monday, 7/28/2014 P09-P12 Even Numbers 11:30 AM-01:30 PM P09-P12 Odd Numbers 4:30 PM — 6:30 PM

Level Accuracy <u>Jeremy Mills</u>, Sagar D. Khare, Jill M. Bolduc, Farhad Forouhar, Vikram K. Mulligan, Scott Lew, Jayaraman Seetharaman, Liang Tong, Barry L. Stoddard, David Baker

POST 03-241 Acrolein-modified High Density
Lipoproteins Promote Atherogenesis Alexandra
Chadwick, Rebecca L. Holme, Yiliang Chen, Kirkwood A.
Pritchard, Daisy Sahoo

POST 03-242 Control of protein production and virus replication by pharmacological blockade of degron detachment Hokyung K. Chung, Conor Jacobs, Yunwen Huo, Jin Yang, Stefanie A. Krumm, Richard K. Plemper, Rodger Y. Tsien, Michael Z. Lin

POST 03-243 Understanding the metabolism of enteric pathogen Campylobacter jejuni Adnan Ayna, Peter Moody

POST 03-244 Chitosan-binding modules (CBM32) of a chitosanse from Paenbacillus sp. IK-5 --- Amino acid residues responsible for chitosan binding--- Shoko Shinya, Takayuki Ohnuma, Hisashi Kimoto, Hideo Kusaoke, Tamo Fukamizo

POST 03-245 Crystal structure of a family GH18 (class V) chitinase from cycad, Cycas revolta ---- structural factors controling the transglycosylation activity of the enzyme Tamo Fukamizo, Naoyuki Umemoto, Takayuki Ohnuma, Toki Taira, Tomoyuki Numata

POST 03-246 Split intein mediated peptide cyclization Shubhendu Palei, Henning D. Mootz

POST 03-247 Exploring the Morpheein Forms of B. cenocepacia HMG-CoA Reductase Riley Peacock, Michelle Brajcich, Courtney Boyd, Jeffrey Watson

POST 03-248 Dynamic Functional Switch in Poliovirus 3C Protease Yan M. Chan, David D. Boehr

POST 03-249 Lighting the Cellular Fuel Gauge with Fluorescent Sensors for Imaging Single-Cell Metabolism Mathew Tantama, Juan Ramón Martínez-François, Rebecca Mongeon, Gary Yellen

POST 03-250 Structural analysis and molecular dynamics of the self-sufficient P450 CYP102A5 and CYP102A1: A combined computational/experimental approach to increase the efficiency of biocatalyst engineering. Maximilian Ebert, Brahm Yachnin, Guillaume Lamoureux, Albert Berghuis, Joelle Pelletier

POST 03-251 Isolation and characterization of proline specific dipeptidyl peptidase IV from the Tenebrio molitor larval midgut Valeriya F. Sharikova, Irina Goptar, Yulia Smirnova, Brenda Oppert, Irina Filippova, Elena Elpidina

POST 03-252 Characterization of hydrolytic enzymeproducing bacteria isolated from paper mill Manel Ghribi, Fatma Meddeb-Mouelhi, Marc Beauregard

POST 03-253 Isolation of NRPS and PKS Gene Clusters from Soil Microbes Danielle N. O'Hara, Connor P. Craig

POST 03-254 Treatment of kraft pulp with enzymes for improving beatability and physical properties
Li Cui, Fatma Meddeb, Marc Beauregard

POST 03-255 Structural and biochemical investigation of the intramolecular interactions of ceramide transfer protein Jennifer Prashek, Seungkyung Kim, Xiaolan Yao

POST 03-256 Evaluating interpretation of B-factors for collective motion modeling <u>Edvin Fuglebakk</u>, Nathalie Reuter, Konrad Hinsen

POST 03-257 Structure/Function Relationships in Carboxylesterase EstGtA2 from Geobacillus thermodenitrificans Jessica K. Moisan, Fatma Meddeb-Mouelhi, Marc Beauregard

POST 03-258 Up-regulation of Rich1 causes S-phase arrest and reduces cell adhesion in epithelial cells Lin Ming-ming, Zhang Qian-ying, Wang Yun-hong, Li Xin, Zhang Jun

POST 03-259 Tracking wood fibers decrystallization with carbohydrate binding module Yannick Hébert-



electrophysiology will be used to study the metabolic components of neurodegeneration in aging and in diseases such as epilepsy and Parkinson's.

POST 03-250

Structural Analysis And Molecular Dynamics Of The Self-Sufficient P450 CYP102A5 And CYP102A1: A Combined Computational/Experimental Approach To Increase The Efficiency Of Biocatalyst Engineering

Maximilian Ebert^{4,1}, Brahm Yachnin², Guillaume Lamoureux^{3,1}, Albert Berghuis^{2,1}, Joelle Pelletier^{4,1} ¹PROTEO, Montreal, Quebec, Canada, ²McGill University, Montreal, Quebec, Canada, ³Concordia University, Montreal, Quebec, Canada, ⁴Université de Montréal, Montreal, Quebec, Canada P450s catalyze the oxidation of non-activated carbon atoms, which is chemically demanding. Members of the CYP102 family are termed "self-sufficient P450s", meaning that they contain all the machinery necessary to ensure the electron transfer and active site regeneration in one single protein. However, the macromolecular assembly remains unknown. Here we report results of SAXS analysis that bring new insights into the formation of the active complex. The recently reported new member CYP102A5 is highly interesting due to its sequence similarity to the intensively studied member CYP102A1, accompanied by a significant increase in electron transfer rate and improved regioselectivity. Homology models were generated, compared and used to identify the structural basis for these differences in catalytic activity. Based on this result we identified residues that may be involved in the gating and substrate capturing mechanism in CYP102A5. Predictions of differences in substrate incorporation and product release from the active site were computed using the adaptive biasing force (ABF) method. With this pioneering application of ABF in enzyme engineering, we were able to predict all known important residues for fatty acid substrate binding in CYP102A1, as well as two additional residues which were identified and analyzed in vitro to support the in silico finding. This newly developed computational biology approach, in addition to conformational studies, will help to guide directed evolution efforts towards the oxidation of non-native substrates.

POST 03-251

Isolation And Characterization Of Proline Specific Dipeptidyl Peptidase IV From The Tenebrio Molitor Larval Midgut

<u>Valeriya F. Sharikova</u>¹, Irina Goptar¹, Yulia Smirnova², Brenda Oppert³, Irina Filippova¹, Elena Elpidina²

¹Chemistry, Lomonosov Moscow State University, Moscow, Moscow, Russian Federation, ²A.N.

Belozersky Institute of Physico-Chemical Biology, Lomonosov Moscow State University, Moscow, Russian Federation, ³Center for Grain and Animal Health Research, Stored Product Insect Research Unit, Manhattan, Kansas, US

The yellow mealworm *Tenebrio molitor* (Coleoptera: Tenebrionidae) is a stored product pest and a biochemical model organism of the family Tenebrionidae. The main food proteins of this insect are storage seed proteins - prolamins, which are also present in the diet of most people. Prolamins contain 10 - 30% proline and 30 - 50% glutamine residues. Several proline- and glutamine-rich prolamin peptides, which are resistant to proteolysis by human digestive enzymes, cause autoimmune Celiac Disease in 1% of the susceptible human population. In this regard, proline specific peptidases (PSP) are urgently needed for therapeutics, capable of hydrolyzing peptide bonds formed by proline residues that are resistant to proteolysis by peptidases of broad substrate specificity. We have carried out a bioinformatic search for PSP in the *T. molitor* larvae gut transcriptome and found 12 sequences similar to human PSP. Two predicted dipeptidyl peptidase IV (DPP IV) sequences were found, one with the highest mRNA expression level among all PSPs in the larval transcriptome. These enzymes are serine exopeptidases from the S9 family, which cleave Xaa-Pro dipeptides from the N-terminus of polypeptides. The major DPP IV-like



enzyme was purified from the *T. molitor* larvae midgut and analyzed by mass-spectrometry analysis. The amino acid sequence of the enzyme coincided with that of the highly-expressed DPP IV found in *T. molitor* gut transcriptome. The isolated enzyme was characterized by substrate specificity, pH-dependence, pH stability, and inhibitor sensitivity. The importance of DPP IV in insect digestion as well as the potential for new treatments of Celiac Disease will be discussed. This work was supported by ISTC grant 3455 and RFBR grants 12-03-01057-a, 14-04-91167-NSFC_a.

POST 03-252

Characterization Of Hydrolytic Enzyme-Producing Bacteria Isolated From Paper Mill

Manel Ghribi^{5, 2}, Fatma Meddeb-Mouelhi^{2, 3}, Marc Beauregard^{1, 2, 4}

¹biologie médicale, Université de Québec à Trois Rivieres, Trois-Rivieres, Quebec, Canada, ²CRML, Trois-Rivieres, Quebec, Canada, ³Buckman North America, Vaudreuil-Dorion, Quebec, Canada, ⁴PROTÉO, Laval, Quebec, Canada, ⁵physique,chimie et biochimie, Université de Québec à Trois-Rivières, Trois-Rivieres, Quebec, Canada

Enzymes act as biocatalyst in many industries, such as textiles, detergent, food, animal feed, bio-fuel, paper and pulp, pharmaceutical, to name a few. Cellulases and hemicellulases are efficient hydrolytic enzymes used in the pulp and paper industry to reduce the cost of production. In addition, industrial enzymes reduce the environmental impact by replacing harmful chemicals. The leading industrial enzyme suppliers (Novozymes, Genencor) offer a limited library of enzymes. Thoses enzymes are poorly adapted to the need of the paper industry. Therefore, our partner, Buckman North America, wants to expand his enzymes portfolio by characterizing new enzymes-producing bacteria. Pulp and paper mills offer untapped biodiversity for microorganisms that use cellulose-based substrates as nutriments. In this project, we will isolate and characterize cellulose and hemicellulose degrading microorganisms from paper mill sludges. To conserve microorganism's biodiversity, we used two temperatures (37 °C and 50 °C) for the isolation step. Detection of extracellular enzymatic activities was carried out on minimum agar plate medium supplemented with either cellulose (carboxylmethylcellulose or Avicel) or beechwood xylans. Bacteria strains showing extracellular cellulase and/or xylanase activities were isolated from various sludges (primary, secondary, presses and machines) found in a paper mill. These bacteria were identified based on their morphology, biochemical characterization and DNA 16s sequencing. The biorefining potential of these enzymes will be evaluated.

POST 03-253

Isolation of NRPS and PKS Gene Clusters from Soil Microbes

Danielle N. O'Hara, Connor P. Craig

University of Richmond, Richmond, Virginia, US

Greater than 90% of microbes living in soil are unculturable due to their complex nutrient and temperature requirements for growth. These microorganisms present a potential source of natural products that could be developed for biotechnological and pharmaceutical uses. Microorganisms with phosphopantetheinyl transferase (PPTase) activity are of high interest due to the role PPTase plays in activating non-ribosomal peptide synthetase (NRPS) and polyketide synthetase (PKS) gene cluster products. The proteins expressed by these gene clusters synthesize complex natural products that are utilized by the microorganism or its host for selective advantages. We screened for NRPS and PKS gene clusters in microbes isolated from soil on the University of Richmond campus. Genomic DNA was isolated from each of these samples and was used to construct metagenomic libraries. We will continue to screen the libraries for PPTase activity in order to identify positive clones. We extended this study to include a library of pigmented microbes previously isolated from Chesapeake Bay sponges, *Clathria*